

Package ‘powerHaDeX’

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Type Package

Title Efficient Simulation of HDX-MS Data and Tools for the
Statistical Analysis

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Description Facilitates simulating and analyzing data coming from HDX-MS
experiments along with the possibility of comparing the power of
the tests verifying differences in the levels of deuterium uptake.
The simulation of mass spectra is a fast version of
<<https://github.com/kanzy/HX-MS-Simulations>>.

License GPL (>= 3)

Encoding UTF-8

Imports checkmate, data.table, expm, ggplot2, glmnet, lme4, lmerTest,
methods, nlme, plyr, Rcpp (>= 1.0.3), signal,

Depends R (>= 3.5.0)

LinkingTo Rcpp

RoxygenNote 7.1.2

Suggests testthat, spelling, knitr, rmarkdown, covr

Language en-US

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powerHDX-package	<i>A short title line describing what the package does</i>
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Description

A more detailed description of what the package does. A length of about one to five lines is recommended.

Details

This section should provide a more detailed overview of how to use the package, including the most important functions.

Author(s)

Your Name, email optional.

Maintainer: Your Name <your@email.com>

References

This optional section can contain literature or other references for background information.

See Also

Optional links to other man pages

Examples

```
## Not run:
## Optional simple examples of the most important functions
## These can be in \dontrun{} and \donttest{} blocks.

## End(Not run)
```

add_column	<i>Complete data frame with columns</i>
------------	---

Description

This function adds column if does not exist and fill it with provided value.

Usage

```
add_column(data, col_name, value = NULL)
```

Arguments

data	a data frame of interest.
col_name	a character. Name of column that should be created if it does not exist
value	optional. A value to fill with.

calculate_hdx_power	<i>Calculate power of statistical tests for HDX experiments</i>
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Description

This function estimates power of statistical tests for HDX experiments.

Usage

```
calculate_hdx_power(
  deuteration_curves,
  tests,
  significance_level = 0.05,
  summarized = TRUE
)
```



```
calculate_hdx_power(deut_curves_p_states,
                   tests = list(test_houde),
                   summarized = TRUE)

n_experiments = 2,
compare_pairs = TRUE,
reference = "all")
```

calculate_peptide_mass

Peptide mass

Description

Calculate mass of undeuterated peptide

Usage

```
calculate_peptide_mass(sequence)
```

Arguments

sequence character vector of amino acid sequence of a peptide

Details

Calculates peptide mass as a sum of amino acids' from sequence masses and H₂O mass (1.007825 * 2 + 15.994915 = 18.01056).

Value

a single number denoting the mass of the undeuterated peptide.

create_experimental_file

Create experimental file

Description

This function generates replications of mass spectra that are consistent with common experimental data files

Usage

```
create_experimental_file(
  peptides,
  times = c(0.167, 1, 5, 25, 1440),
  charge,
  n_replicates = 3,
  mass_deviations = 50,
  intensity_deviations = NULL,
  file_type = "DynamX"
)
```

Arguments

peptides	a data frame of sequences (sequence), Protein, and Start, End and parameters except times that can be used for simulating mass spectra. See simulate_theoretical_spectra for more details about the additional parameters.
times	a vector of times at which deuteration levels will be measured (seconds)
charge	vector of charges of the peptide ion. If NULL, one value is sampled from vector 2:6. Default NULL.
n_replicates	number of technical replicates to create
mass_deviations	mass deviation in parts per million. Either a single number (then the error at each time point will be the same) or a vector of the same length as number of unique time points in the experiment. The error will be sampled from normal distribution with standard deviation equal to $mass_deviations * undeuterated_{mass}/1e6$ Default to 50.
intensity_deviations	optional, standard deviations of random noise that will be added to intensities. Either a single number (then the error at each time point will be the same) or a vector of the same length as number of unique time points in the experiment. The error will be sampled from normal distribution with these standard deviations. Default NULL.
file_type	the type of file. Default to "DynamX".

Value

data table. The table of HDX-MS results consistent with 'file_type' format.

Examples

```
peptides <- data.frame(sequence = c("FPTTKTY", "LVRKDLQN"),
  protection_factor = c(10, 100))
create_experimental_file(peptides, charge = 1:3)
```

`fix_columns_names_types`*Standardize column names and types*

Description

Standardize column names and types

Usage

```
fix_columns_names_types(queries)
```

Arguments

queries list of lists of data.tables

`get_deuteration_single_timepoint`*Calculates deuteration for given timepoint*

Description

Calculates deuteration for given timepoint

Usage

```
get_deuteration_single_timepoint(  
  initial_matrix,  
  time_sequence,  
  hd_probs,  
  dh_probs  
)
```

Arguments

initial_matrix A matrix
time_sequence vector of exchange times
hd_probs probabilities of transition HD
dh_probs probabilities of transition DH

Value

a matrix denoting hydrogen-deuterium exchange for given timepoint.

```
get_noisy_deuteration_curves
```

Replicated deuterium uptake curves

Description

This function creates a list of lists of noisy deuteration curves based on theoretical spectra in order to imitate the data from the HDX experiments.

Usage

```
get_noisy_deuteration_curves(  
  theoretical_spectra,  
  compare_pairs = TRUE,  
  reference = NA,  
  n_replicates = 4,  
  n_experiments = 100,  
  mass_deviations = 50,  
  intensity_deviations = NULL,  
  per_run_deviations = NULL,  
  relative = TRUE  
)
```

Arguments

theoretical_spectra	a data table or a list of data tables of theoretical spectra created by the function simulate_theoretical_spectra .
compare_pairs	if FALSE, all groups (defined by the protection factor) will be considered jointly. If TRUE (default), each protection factor will be considered together with the protection factor given by the 'reference' parameter.
reference	protection factor that will be used for comparison to other protection factors in. The function accepts either NA (for comparing all protection factors), a number (for comparing with reference value of protection factor) or "all" (for pairwise comparisons of all the possible combinations). Default NA.
n_replicates	number of technical replicates to create
n_experiments	number of replicates of an experiment for power calculation.
mass_deviations	mass deviation in parts per million. Either a single number (then the error at each time point will be the same) or a vector of the same length as number of unique time points in the experiment. The error will be sampled from normal distribution with standard deviation equal to

$$mass_deviations * undeuterated_mass / 1e6$$

Default to 50.


```

use_markov = TRUE)

theo_spectra_two_states <- rbind(theo_spectra_pf_100, theo_spectra_pf_200)

deut_curves_p_states <- get_noisy_deuteration_curves(theo_spectra_two_states,
                                                    n_replicates = 4,
                                                    n_experiments = 2,
                                                    compare_pairs = TRUE,
                                                    reference = "all")

```

get_spectra_list *Get a list of spectra*

Description

Create a list of data tables of spectra for all states jointly or paired states.

Usage

```
get_spectra_list(theoretical_spectra, compare_pairs = FALSE, reference = NA)
```

Arguments

theoretical_spectra	a data table or a list of data tables of theoretical spectra created by the function simulate_theoretical_spectra .
compare_pairs	if FALSE, all groups (defined by the protection factor) will be considered jointly. If TRUE (default), each protection factor will be considered together with the protection factor given by the 'reference' parameter.
reference	protection factor that will be used for comparison to other protection factors in. The function accepts either NA (for comparing all protection factors), a number (for comparing with reference value of protection factor) or "all" (for pairwise comparisons of all the possible combinations). Default NA.

Details

If the parameter `compare_pairs` is FALSE then all the provided protection factors will be considered jointly. If `compare_pairs` is TRUE, then the parameter `reference` is necessary (a single number or "all"). Then the data is split via the supplementary function [get_paired_spectra](#) into data tables of spectra with paired biological states (the reference protection factor and the protection factor of interest if provided, or all the possible pairs if reference equals "all").

Value

list of data.tables containing spectra - for paired states or all states.


```
plot_spectra(theo_spectra)
```

powerHaDeX

powerHaDeX

Description

The R-package powerHaDeX for simulating and analyzing data coming from HDX-MS experiments along with the possibility of comparing the power of the tests verifying differences in deuteration levels.

Author(s)

Michal Burdukiewicz, Krystyna Grzesiak, Mateusz Staniak

prepare_input_peptides

Prepare input for [create_experimental_file](#)

Description

Supplementary function providing appropriate input.

Usage

```
prepare_input_peptides(peptides)
```

Arguments

peptides a data frame of parameters for which [simulate_theoretical_spectra](#) will be executed.

Value

a data frame being a proper input for [create_experimental_file](#).

 simulate_theoretical_spectra

Simulate theoretical spectra of a deuterated peptide over time

Description

Simulate theoretical spectra of a deuterated peptide over time

Usage

```
simulate_theoretical_spectra(
  sequence,
  charge = NULL,
  protection_factor = 1,
  times = c(60, 600),
  pH = 7.5,
  temperature = 15,
  n_molecules = 100,
  time_step_const = 1,
  if_corr = FALSE,
  min_probability = 1e-04,
  use_markov = TRUE
)
```

Arguments

sequence	amino acid sequence of a peptide as a single string
charge	vector of charges of the peptide ion. If NULL, one value is sampled from vector 2:6. Default NULL.
protection_factor	protection factor. If a single number of provided, same protection factor will be assumed for each amide. Default value: 1 (indicates that the exchange rate is equal to the intrinsic exchange rate)
times	a vector of times at which deuteration levels will be measured (seconds)
pH	pH of the reaction. Default to 7.5.
temperature	temperature of the reaction (Celsius)
n_molecules	number of peptide molecules. Default to 100.
time_step_const	time step constant. Default value: 1. Value that indicates the length of the time step of the simulation. The bigger the time step, the fewer time points are simulated (the fewer iterations in case of Zhong-Yuan Kan's approach).
if_corr	logical. PH correction indicator. Default value FALSE. The value of pH is equal to pD. If there is correction, the pD = pH + 0.4. (Connelly et al 1993)
min_probability	smallest isotopic probability to consider

`use_markov` logical. If TRUE algorithm basing on Markov chain will be used. If FALSE simulation provided by Zhong-Yuan Kan will be executed. Default to TRUE, as it fastens the calculation

Details

To the results calculated by `get_iso_probs_deut` is added a minimal exchange control - for time point 0 (directly after adding a buffer). The m/z values are obtained as a ratio of the `peptide_mass` magnified by proton mass and the peptide charge. The distribution of undeuterated peptide is the intensities vector.

Value

a data table of variables:

- Exposure - time point of a measurement,
- Mz - mass-to-charge ratio,
- Intensity - isotopic probabilities larger than `min_probability`(the smaller ones are zeroes)

and the variables provided by user

- Sequence,
- PF,
- Charge,
- PH.

See Also

The algorithm that is used to simulate theoretical spectra is based on Zhong-Yuan Kan's implementation in Matlab. The original version of codes is located in the repository <https://github.com/kanzy/HX-MS-Simulations> (as at 29.06.2020). In the powerHaDeX package can be found the Kan's algorithm re-implemented in R (using Rcpp) and the accelerated implementation (that uses Markov chains' properties). Moreover, the package powerHaDeX allows the user to simulate spectra for more than one exposure time for both (Rcpp and Markov) approaches.

Examples

```
simulate_theoretical_spectra(sequence = "LVRKDLQN",
                             charge = c(3, 5),
                             protection_factor = 100,
                             times = c(0.167, 5),
                             pH = 7.5,
                             temperature = 15,
                             n_molecules = 500,
                             time_step_const = 1,
                             use_markov = TRUE)
```

test_hadex_data	<i>Apply tests for HaDeX data</i>
-----------------	-----------------------------------

Description

This function converts the data from HaDeX in order to make it compatible with the input of test functions and perform the testing procedures of provided tests.

Usage

```
test_hadex_data(  
  dat,  
  states = unique(dat[["State"]])[1:2],  
  tests = list(test_houde)  
)
```

Arguments

dat	data.table. The data of hdx_data class from the HaDeX package.
states	a character vector containing two states from provided 'dat' that should be tested. By default the first two states (if exist) from 'dat' are chosen.
tests	a list of testing functions. In the 'powerHaDeX' package the following tests are implemented: - test_houde , - test_hdx_analyzer , - test_memhdx_model , - test_semiparametric .

Value

This function returns a data table of variables:

- Test - name of test,
- State_1, State_2 - tested states from states,
- Significant_difference - TRUE or FALSE, indicating whether the null hypothesis is rejected
- Sequence - amino acid sequence that was tested


```
theo_spectra_pf_200 <- simulate_theoretical_spectra(sequence = "LVRKDLQN",
                                                    charge = c(3, 5),
                                                    protection_factor = 200,
                                                    times = c(0.167, 5),
                                                    pH = 7.5,
                                                    temperature = 15,
                                                    n_molecules = 500,
                                                    time_step_const = 1,
                                                    use_markov = TRUE)

theo_spectra_two_states <- rbind(theo_spectra_pf_100, theo_spectra_pf_200)

deut_curves_p_states <- get_noisy_deuteration_curves(theo_spectra_two_states,
                                                    n_replicates = 4,
                                                    n_experiments = 1,
                                                    reference = 100)[[1]][[1]]

test_hdx_analyzer(deut_curves_p_states)
```

test_houde

Houde's test for deuteration curves

Description

This function performs Damian Houde's confidence intervals test for differences in deuteration levels. Its input and output are compatible with the function [calculate_hdx_power](#).

Usage

```
test_houde(data, significance_level = 0.05)
```

Arguments

`data` data.table with deuteration curves
`significance_level` significance level for tests

Value

This function returns a data table compatible with the function [calculate_hdx_power](#).

References

Houde, Damian, Steven A Berkowitz, and John R Engen (2011). "The utility of hydrogen/deuterium exchange mass spectrometry in biopharmaceutical comparability studies". In: *Journal of pharmaceutical sciences* 100.6, pp. 2071–2086.

See Also

Other tests:

- [test_hdx_analyzer](#)
- [test_memhdx_model](#)
- [test_semiparametric](#)

Or [calculate_hdx_power](#) for estimation of power of tests for differences in deuteration levels.

Examples

```
theo_spectra_pf_100 <- simulate_theoretical_spectra(sequence = "LVRKDLQN",
                                                    charge = c(3, 5),
                                                    protection_factor = 100,
                                                    times = c(0.167, 5),
                                                    pH = 7.5,
                                                    temperature = 15,
                                                    n_molecules = 500,
                                                    time_step_const = 1,
                                                    use_markov = TRUE)
theo_spectra_pf_200 <- simulate_theoretical_spectra(sequence = "LVRKDLQN",
                                                    charge = c(3, 5),
                                                    protection_factor = 200,
                                                    times = c(0.167, 5),
                                                    pH = 7.5,
                                                    temperature = 15,
                                                    n_molecules = 500,
                                                    time_step_const = 1,
                                                    use_markov = TRUE)

theo_spectra_two_states <- rbind(theo_spectra_pf_100, theo_spectra_pf_200)

deut_curves_p_states <- get_noisy_deuteration_curves(theo_spectra_two_states,
                                                    n_replicates = 4,
                                                    n_experiments = 1,
                                                    reference = 100)[[1]][[1]]

test_houde(deut_curves_p_states)
```

test_memhdx_model *MEMHDX model*

Description

This function performs the test based on a linear mixed effects model used in MEMHDX tools. Its input and output are compatible with the function [calculate_hdx_power](#).

Usage

```
test_memhdx_model(data, significance_level = 0.05)
```



```
time_step_const = 1,
use_markov = TRUE)
theo_spectra_pf_200 <- simulate_theoretical_spectra(sequence = "LVRKDLQN",
charge = c(3, 5),
protection_factor = 200,
times = c(0.167, 5, 10, 30),
pH = 7.5,
temperature = 15,
n_molecules = 500,
time_step_const = 1,
use_markov = TRUE)

theo_spectra_two_states <- rbind(theo_spectra_pf_100, theo_spectra_pf_200)

deut_curves_p_states <- get_noisy_deuteration_curves(theo_spectra_two_states,
n_replicates = 4,
n_experiments = 1,
reference = 100)[[1]][[1]]

test_semiparametric(deut_curves_p_states)
```

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